Cheatography

Micrometry Technique Cheat Sheet by UmeshJagtap via cheatography.com/186232/cs/39996/

AIM

Measurements using the Ocular Micrometer (Micrometry Technique).

OBJECTIVES

- 1. Calibrate the Ocular Micrometer Scale.
- 2. Measure samples using the Ocular Micrometer Scale.

INTRODUCTION

Micrometer scales within the ocular of a microscope are employed to determine sample sizes. These specialized oculars possess a transparent scale graticule superimposed onto the observed image. By replacing the stage micrometer with the sample slide, you can measure cell sizes. Micrometry is a crucial technique in biology, especially in microscopy, using ocular and stage micrometers to measure biological structures.

1. OCULAR MICROMETER

Descri- ption:	Transparent ruler-like device without units, mounted on eyepiece.					
Design:	Glass disc featuring 10 mm scale divided into 100 sub- divisions.					
Visibility:	Scale visible through microscope eyepiece.					
2. STAGE MICROMETER						
Placement:	Positioned on microscope stage.					
Constr-	Slide with 2 mm scale divided into 0.01 mm (10 $\mu\text{m})$					

Function: Used to calculate ocular micrometer divisions at specific magnifications.

REQUIREMENTS

uction:

- Compound light microscope with bjectives (4X, 10X, 20X, 40X),
- Ocular micrometer scale
- □ Stage micrometer (0.01 mm scale).

sub-divisions

- □ Microscope Glass slide
- □ Coverslip

PROTOCOL

Part I: Calibration of Ocular Micrometer

- 1. Place the ocular micrometer in the microscope eyepiece.
- 2. Adjust the lighting for optimal Kohler illumination.
- 3. Focus on the stage micrometer using the lowest magnification.
- 4. Rotate to the objective designated for calibration.

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5.Adjust the focus of the ocular micrometer to visualize both scales

PROTOCOL (cont)

clearly.

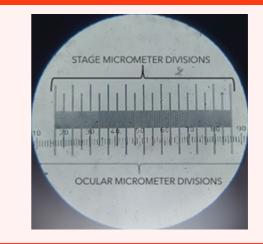
6. Align the stage and ocular scales with a slight offset for ease of reading.

7. Choose two positions on each scale, preferably on opposite sides.

8. Count the divisions on the stage scale (ssd) between the chosen positions.

- 9. Count the divisions on the ocular scale (osd), which equals ssd.
- 10. Record the number of ocular spaces (y) and stage spaces (x).
- 11.Calculate the calibration factor: 1 ocular space = (x/y) \times 10 $\mu m.$
- 12. Repeat the process for the second assigned objective.

Figure1:



Microscope Field of View Showing Ocular Micrometer and Stage Micrometer Superimposed each other.

Observation Table 1: Calibration of Objectives								
Objective Power	Obser vation No.	Ocular Divisions (Y)	Stage Divisions (X)	Calibration Factor (X/Y × 10 µm)	Mean Calibr- ation factor			
10X	1 2							
	3							
40X	1							
	3							

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Part II: Protocol for Using Micrometers

- 1. Place biological sample slide (e.g. Pollen Grain), focus on area.
- 2. Locate ocular micrometer through eyepiece.
- 3. Align structure with ocular micrometer.
- 4. Count ocular divisions needed to span the structure.
- 5. Repeat for other objectives and other biological specimens.
- 6. Record divisions for each measurement.
- 7. Calculate structure size (e.g. pollen grain): Ocular divisions × calibration factor.
- 8. This protocol accurately measures biological structures.

Observation Table2 :Measurement of Plant Samples

Plant Samples	Microscope Objective Power	Calibr ation Factor (CF)	Obser vation No.	Ocular Divisions (OSD)	Size of Sample (OSD × CF)	Mean Size (µm)	
Pollen Grain	10X		1				
			2				
			3				
	40X		1				
			2				
			3				

RESULTS

Size of Pollen Grain under 10X =	μm
Size of Pollen Grain under 40X =	μm

CONCLUSION

In conclusion, micrometer scales within microscope oculars are pivotal for accurate sample size determination in biology. By replacing the stage micrometer with a sample slide, cell sizes can be measured



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