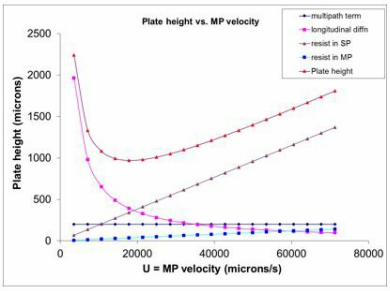
Basic Theory

Gas Chromatography	SP → Liquid
	MP→ Inert gas ▸ No role in separation ▸ Only directs analyte down column (carrier gas)
	$D_m >>> D_s$ ▸ $C_mU \sim 0$
	Flow rate ▸ Dictate by choice of SP (thickness, properties) ▸ Modest plate height ~1mm (↑ L = ↑N)

Theory Equations



$$A = 2\lambda d_p \qquad \frac{B}{U} = \frac{2\gamma D_m}{U}$$

$$C_sU = \frac{f_s(k')d_f^2}{D_s}U$$

$$C_mU = \frac{f_m(k')d_p^2}{D_m}U \approx 0$$

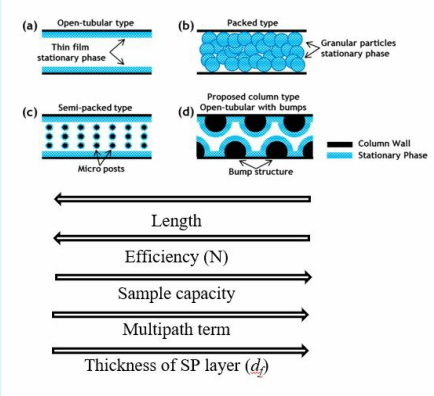
Column Type

Packed	Packed full of particles
	Put SP on particles
	MP pushes through packed bed
	Tubing ▸ Glass, stainless steel, etc. ▸ Inert = not part of separation

Column Type (cont)

Wall Coated Open Tubular (WCOT)	Inside wall of quartz/glass tube ▸ Chemically roughen ▸ ↑ Surface area ▸ Coated with SP
Fused Silica Open Tubular (FSOT)	SP coating on wall of long thin tube ▸ Smooth wall
	Diameter ~ 75-200 um

Column Diagram



GC Systems

Sample	Introduce into injector port ▸ Vaporize sample ▸ Vaporized analyte swept into column
Mobile phase	High pressure cylinders ▸ Use a gas flow regulator ▸ Regulate the pressure
Detector	Detect components of the mixture being eluted off the chromatography column ▸ Some may require a reference flow
Oven	Separation occurs ▸ Controlled temperature ▸ Fan → Circulates air and controls temperature

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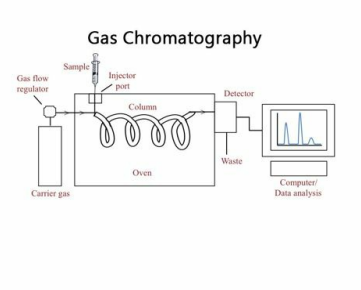
Cheatography

Gas Chromatography Cheat Sheet

by shaylannxd via cheatography.com/149855/cs/32547/

GC System Diagram



Split Flow Injector

Sample dissolved in volatile solvent

Collect sample into syringe and inject through rubber septum

- Seals injector for analyte to go into column
- Protects from outside atmosphere
- Bad peak shapes = hole in septum

Use a heat block to "flash" sample into vapour

- ~50-100C hotter than oven
- Need to vaporize sample

GC systems design to operate with 3 main columns

- Each column has a different flow rate
- Adjust based on column used

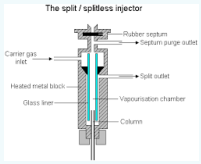
FSOT/WCOT

- Can't handle large sample mass
- Small diameter
- Limited SP
- Limited volume capacity
- Control by valve system

Split flow outlet

- Avoid overloading the column
- Packed → Set at 0 (closed)
- FSOT/WCOT → Split flow ratio → Depends on [analyte] in injection volume

Split Flow



Requirements of SP

Requirements of SP (cont)

	Prefer a low volatile solvent → Don't want SP to vaporize in oven
Thermal Stability	SP in column
	Don't want thermal breakdown products
Inert/Reactive	Don't want analyte to react with SP
	Only want to interact

Stationary Phase

Siloxane Polymer	Low volatility
	Thermally stable bond
	Contains a silicone backbone ▸ Close to inert ▸ Can be derivatized
	Add pendant functional groups ▸ Tune selectivity/solubility/retention ▸ Adjust polarity
Non-Polar	Poly(dimethyl)siloxane (PDMS) ▸ Good quality
	Flurocarbons
Polar	Can replace dimethyl/methyl groups ▸ CN,CO,OH
Phenyl Groups (Benzene Ring)	Non-polar
	π e^- → delocalized
	When approach by polar molecules ▸ e^- reorganized → Induced dipole interactions ▸ Can behave polar with polar molecules (vice versa)
Chiral Moiety	Chiral-chiral interactions on SP
	Rise to selectivity of 1 enantiomer over another

Solvent	Must dissolve analyte
	Bad SP ▸ Unretained ▸ Affects R'~0 ▸ No separation
Volatility	A substance with high volatility is more likely to exist as a vapour
	A substance with low volatility is more likely to be a liquid or solid

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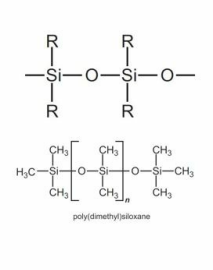
Siloxane Polymer



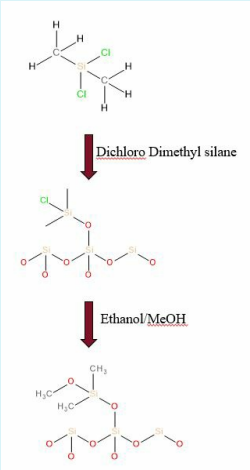
Minimize Loss of SP

Bonded Phase	Process of the SP polymer is attached to ▸ Silica support particle ▸ Wall of a capillary
	A liquid-liquid chromatography method in which a stationary phase is covalently bound to a carrier particle
Cross--Linked Phase	Polymer attached to wall
	Polymer cross-linked with each other ▸ Critical for separation
	Produce more rigidty, hardness and ↑ Melting point ▸ Formation of covalent bonds
Issue	Most SP are non-polar and silica support surface are polar ▸ Not much intertaction
	Uses phases to prevent issue of contact
	Use silane reaction to bond/cross-link
Silane Reaction	Use to anchor/bond silicones to silica surfaces ▸ In packing materials (particles) ▸ FS capillaries
	Use to deactivate silanols
	Same chemistry for polymerization and cross-linking

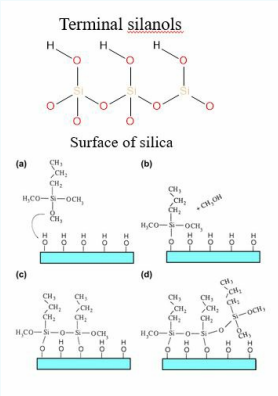
Minimize Loss of SP (cont)

	Silanol ▸ Very polar ▸ Expose on surface of silica ▸ Disagree with SP polarity ▸ Competition for polar analyte
	Deactivation chemistry ▸ Use dichloro dimethyl silane ▸ Use ethanol/MeOH ▸ Create less polar surface
Inertness of Column	Residual silanols ▸ React strongly to polar compounds ▸ Produce tailing peaks ▸ Undesirable interactions in column

Deactivation of Silanol



Silane Reaction Mechanism



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Controlling Retention

Retention	Controls resolving power (R') ▸ R' depends on K' ▸ K' depends on separation conditions
	Want all peaks to fall in "ideal range" of retention ▸ 1-10
	MP is inert ▸ Only function to control retention ▸ Equilibrium constant = thermodynamic property
	Temperature ▸ Alter overall retention
	Type of SP ▸ Alter selectivity
Impact of Different Temperature	Isothermal separation ▸ A thermodynamic process, in which the temperature of the system remains constan
	↓ Temperature = ↓ Thermal energy available ▸ Less thermal energy ▸ Analyte spends more time in SP ▸ More time in column ▸ Clearer separation
	↓ Temperature =↑ Resolution = ↑ Overall time ▸ Can become excessive ▸ Needs to adjust separation as it proceeds
	↓ Temperature → Favors SP
	↑ Temperature → Favors MP
Different Ramp Rates	Altered t_r and resolution independently

Controlling Retention (cont)

	Adjust temperature during course of separation
	Resolution imporves under better retention conditions for the analyte
	Change in gradient steep → Improves separation ▸ Shorten separation time ▸ Increase resolution ▸ As a function of temperature
Round-Up of T Programming	Powerful tool for controlling K'
	Directly affects distribution constant
	↑ Temperature = ↓ K'
	Ramped (gradient) temperature is used to adjust K'
	Make GC less intuitive ▸ ↓ R= {{fa-arrow down}} K' (general) ▸ ↑ R= ↓ K' (T programming) ▸ K'=f(T)
	Separation limited by ΔT/Δt (ramp rate)
	Column lifetime is shorter at higher temperature
Other Factors	$K'=K(V_s/V_m)$ ▸ SP thickness ▸ Total mass of SP
	FSOT columns ▸ Calculate phase ratio (V_s/V_m)

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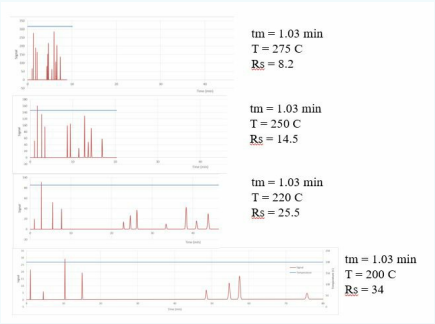
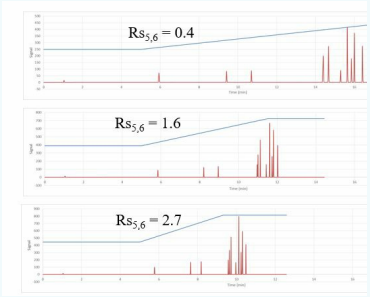
Graphs with Different Temperature



Ramp Rate Graphs



GC Detectors

Requirements

Sensitivity
- 10^{-8}-10^{-15} g analyte/s
- Packed → All sample used → Decrease efficiency = broader peaks
- FSOT → Split flow injector (5-10% sample used)→ Increase efficiency = narrow peaks

Stability
- Noise on baseline → Smooth {[fa-arrow-right]} Detect the smallest peaks → Minimal DL
- Drift → No baseline (goes up and down)

LDR
- 5-8 orders of magnitude

GC Detectors (cont)

Can accept MP over a wide temperature range
- T Programming {[fa-arrow-right]} Improves separation
- Immune to T change
- Compensate T change → Require reference gas flow

Fast response and independent of T

Simple to use, maintain, repair

Selective/Universal
- Detect analyte of interest (S)
- Detect all species (U)

Non-destructive

Flame Ionization Detector (FID)

Analyte elute from column
- Mix with H2 gas
- Combusted

Reduced carbons
- Produce ions that alter conductivity of flame and alter current

Signal proportional to # of reduced carbons
- Mass sensitive

Oxidized and e^- capturing species
- No-little signal
- Cannot be oxidized further

Non-combustible gasses
- No signal
- Already oxidized

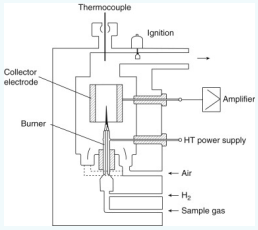
High sensitivity
- 10^-13& g/s
- use FSOT/WCOT

Large LDR
- 7 orders of magnitude

Destructive

No reference flow

FID Diagram



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Electron Capturing Detector (ECD)

An ionization detector
- response is based upon the ability of molecules with certain functional groups to capture electrons generated by the radioactive source

Radioactive source → ^{63}Ni
- Emits beta-particles

When disintegration occurs
- Large energy release
- Beta particle emission
- Impacts any filler gas and/or MP present in detector and ionize it

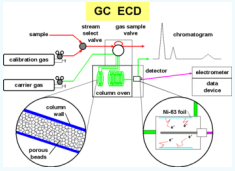
Use a N2 make-up gas
- Get ionized by high energy
- Ionized N2 gas → Pass an electric current through detector cell

In absence of analyte with e^- capturing groups
- A constant current established through the detector

When analyte with e^- capturing groups enters cell
- Quench some ionization
- Reduce conductivity of gas = reduce current in cell

Selective detector
- analytes with a high electron affinity
- Sensitive for species that can disrupt ionization of N2 gas
- Pesticides →halides, peroxides, nitro groups

ECD Diagram



Thermal Conductivity Detector(TCD)

Properties	Signal proportional to change in heat capacity ▸ Difference between MP and MP+analyte are relatively small
	Universal detector ▸ Detect solvent as well ▸ Undersirable → Solvent order of magnitude is more concentrated than analyte ▸ Result in large solvent peaks and small analyte peaks ▸ If analyte is not well retained → Interfered by solvent
	Modest sensitivity ~ $10^{-9 \text{ to } -10}$ g/ml ▸ Less sensitive than FID
	Modest LDR ▸ Very short linearity
	Non-destructive
	Require a reference flow
Basic Theory	Based on ability of the gas exciting the column to absorb heat
	Contains thin filament ▸ electrically heated ▸ As heat capacity of gas changes (MP vs MP+analyte), so does the T of the filament
	Resistance of thin filament ▸ T changes the resistance ▸ Resistance changes the current of the circuit →Current is VERY sensitive to T
Reference Flow (Type 1)	To compensate for the T of MP coming from the oven ▸ T is changing with T programmed elution ▸ left section of diagram
	Equation ▸ $V_{out1} = V_{applied} * (R_{ref}/(R_{column}+R_{ref}))$
	If $R_{column} = R_{ref}$ ▸ two resistors are “balanced” →The signal from the column is coming from the MP ▸ $V_{out1}=(1/2)V_{app}$

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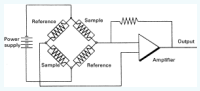
Thermal Conductivity Detector(TCD) (cont)

	If $R_{column} \neq R_{ref}$ ▸ Analyte's heat capacity changes T ➔heating or cooling of filament ▸ V_{out1} increases as analyte elutes ➔As R_{column} gets closer to 0, V_{out1} gets closer to V_{app}
Reference Flow (Type 2)	Opposite concept as reference flow type 1 ▸ right section of diagram
	Equation ▸ $V_{out2} = V_{applied} * (R_{column}/(R_{column}+R_{ref}))$
	If $R_{column} = R_{ref}$ ▸ two resistors are "balanced" ➔The signal from the column is coming from the MP ▸ $V_{out1}=(1/2)V_{app}$
	If $R_{column} \neq R_{ref}$ ▸ Analyte's heat capacity changes T ➔Heating or cooling of filament ▸ V_{out2} decreases as analyte elutes ➔ As R_{column} gets closer to 0, V_{out2} gets closer to 0
Reference Flow (Type 3)	Type 1 and Type 2 TCD operating together ▸ With separate power supplies
	If $R_{column} = R_{ref}$ ▸ two resistors are "balanced" ➔The signal from the column is coming from the MP ▸ $V_{out1}=(1/2)V_{app}$

Thermal Conductivity Detector(TCD) (cont)

	If Rcolumn ≠ Rref ▸ As analyte elutes ➔V_{out1} increases ➔V_{out2} decreases ▸ V_{out1} and V_{out2} have same magnitude, opposite signs ➔Taking the difference of the two will double the V measured ➔Double the signal for the same effort
Reference Flow (Type 4)	Same as Type 3, but with a single power supply
	Wheatstone bridge ▸ Name of this circuit ▸ Common approach for detecting VERY small change in resistance ➔Advantage: Doubles the signal magnitude

TCD Diagram



GC-MS

Properties	Versatile ▸ Provide identification power
	Have to run known standards (spiked)
	Electron beam ionization ▸ M^+ and fragments
	Excellent DL ▸ Depending on instrument and analyte ▸ ~ 2-20 picog injected
	Concentration DL in sample ▸ Depends on sample work-up
	Long LDR ▸ Dependent on instrument ▸ 4-6 orders of magnitude

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GC-MS (cont)

Selective
- Less interferences
- Filters out MP signal

Destructive

Expensive

Basic Theory

Quadruple MS
- Contains 2 positive and 2 negative poles

Movement of M^+
- M^+ travels in a sinusoidal path
- If M^+ is too light or too heavy, it is kicked out of quadrupole

→b/c they are not really able to respond to polarity change
- How to fix this

→Quickly change the frequency and voltage of the poles

→Can quickly scan through all m/z ratio to obtain mass spectrum

Spectrum generated
- Total ion current (TIC)

→Easiest way

→Sum of all ion signals that passes through

→Acts as a universal detector: does not filter out MP signal

→Tells you how many species are present
- Extracted mass spectra

→Take a slice of TIC peak and see its fragments

GC-MS (cont)

Isotopes
- Parent ion

→ Most prominent and heaviest
- Isotopes

→Daughter peak from most prominent peaks

→can provide more info depending on its ratio with parent peak
- Isotopically labelled analytes

→Replacing parts of molecule with deuterium

→Produces a known mass higher than the original mass

→Compare spectrum with orignal

Positive identification
- Compare experimental spectrum with the “real” analyte spectrum
- 3 steps

→Correct mass of molecule?

→Correct set of fragments?

→Correct fragment intensities?

Quantitation
- Usually multiple ions monitored/measured

→Validate ratio of peaks at the correct m/z ratio

Column bleed
- SP is boiling and bleeding out
- Leads to a rise in baseline

→Not good
- How to fix it
- Running at low T

→Purchase column made specifically for MS ($$$)

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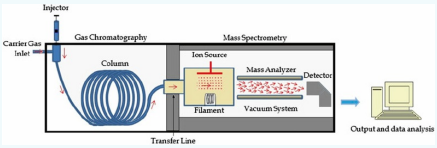
GC-MS Diagram



Key Factors and Applications

When will GC be useful	1. Analyte ▸ Needs to be volatile ▸ Not proteins → Unstable at high temperature ▸ Silation reaction → Produce volatile products (Risk of contamination, loss, produce new products) ▸ Needs to be stable → Stable enough to transit the column
	2. High enough concentration to detect ▸ Packed columns: great sample capacity but low resolving power and resolution ▸ FSOT: lower capacity (split flow) but high resolving power and resolution ▸ Detectors: Has a good sensitivity
	3. Does sample require high R' separation ▸ Depends on the type of detector ▸ Universal = high R ▸ Selective = low R
	4. Generally faster than LC
Applic-ations	Anti-dopping and forensics
	BAC (Crime/forensics labs)
	Pharmaceuticals ▸ Process control ▸ Quality control ▸ Research and development

Key Factors and Applications (cont)

Food and Beverages
- ▸ Wine/alcohol
- ▸ Pesticides

Environmental
- ▸ Pesticides
- ▸ PAH and industrial solvents
- ▸ Oil/hydrocarbon spills

R&D
- ▸ Organic synthesis
- ▸ Catalysis (monitor products)

Industrial
- ▸ Feedstock
- ▸ Off gassing

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